

**Next Generation Sequencing –
The Role of New Sequence Technologies in Shaping the
Future of Veterinary Science**

Hosted by the RCVS Charitable Trust



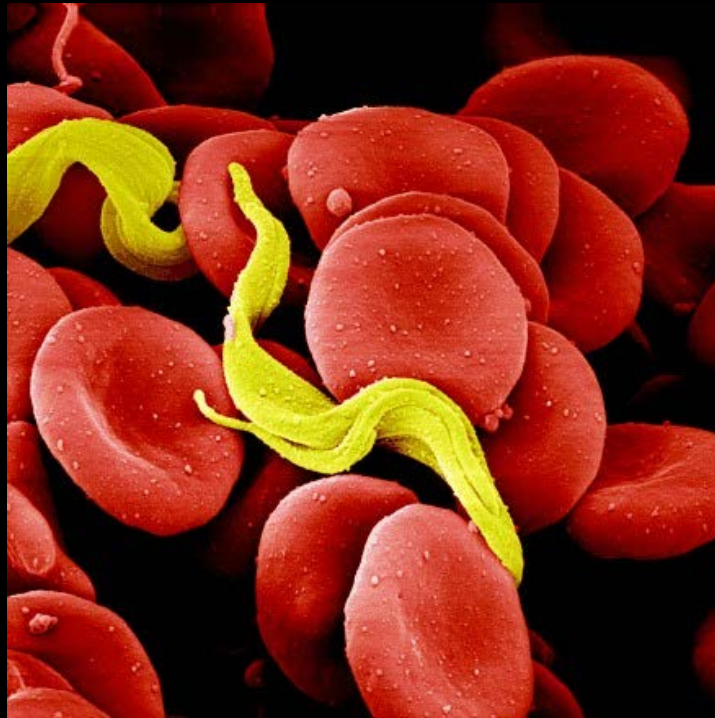


UNIVERSITY OF
LIVERPOOL

Sequencing pathogens (and their hosts): from large to small, from virulent to benign

Trypanosomiasis

- **“Sleeping Sickness” in humans**
 - 500,000 affected annually
 - 37 countries; 22 of which least developed in the world
 - Fatal if untreated
- **“Nagana” in cattle**
 - Roughly 1/3 of African cattle population ‘at risk’
 - Causes cachexia, anaemia and increased susceptibility to secondary disease
 - Losses in livestock and crop production amount to an estimated \$5 billion annually



Trypanosomiasis: “Sleeping Sickness” – Human Disease

Affects 500,000 annually across 36 countries

Caused by two species of *T. brucei* parasite:

T. brucei gambiense

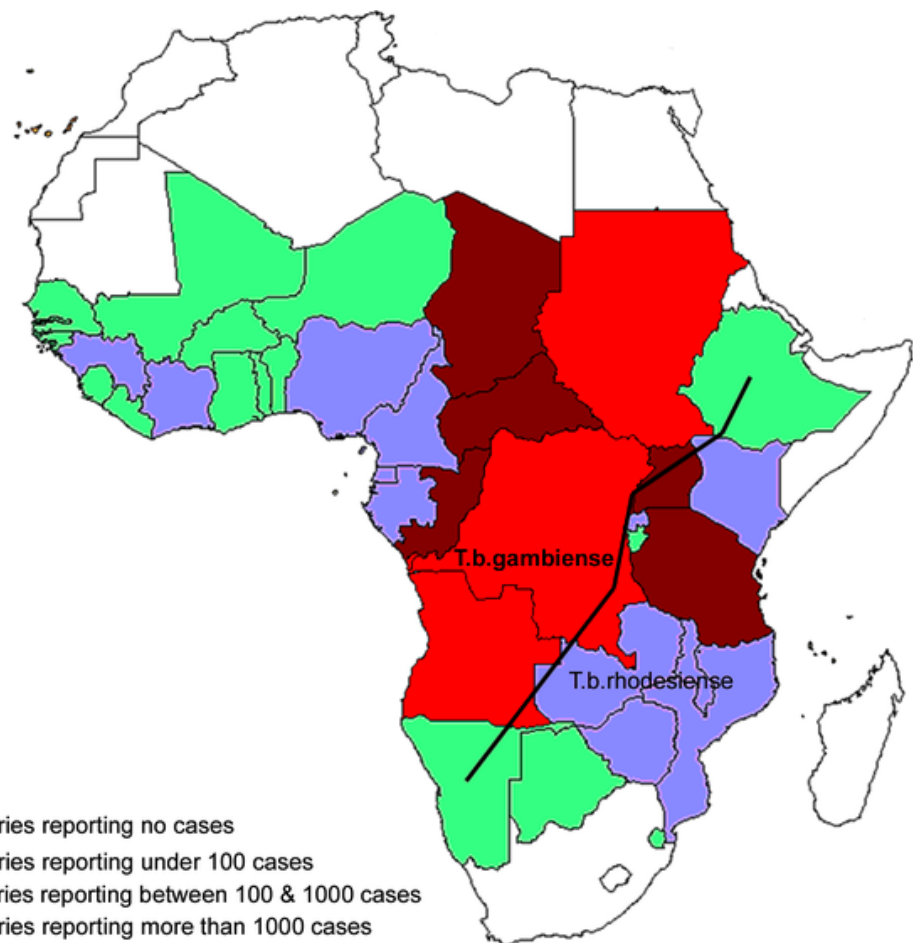
- Western Africa
- Chronic disease

T. brucei rhodesiense

- Eastern Africa
- Acute Disease

T. brucei brucei

- widespread
- disease of cattle and other non-human vertebrates

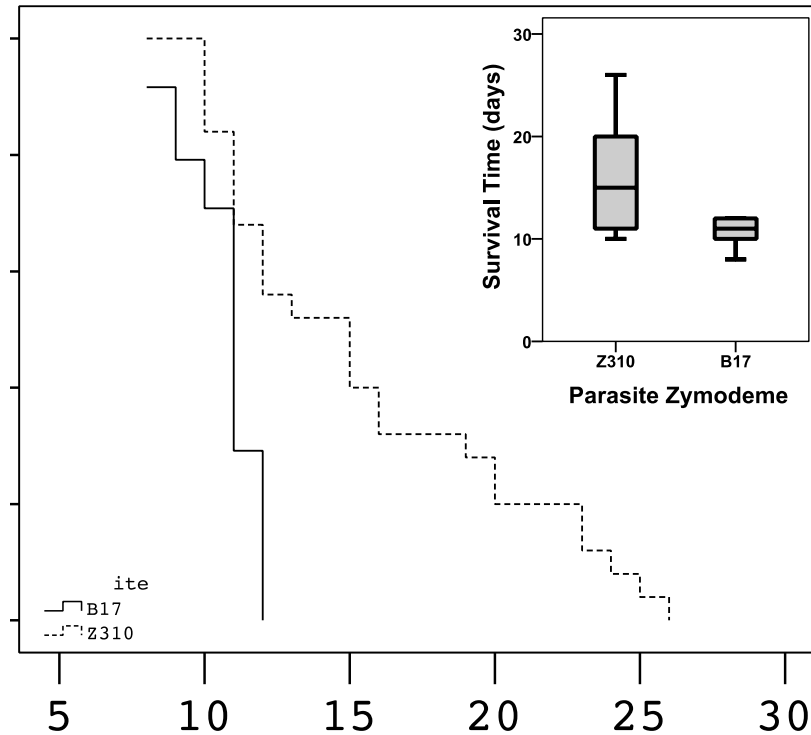


Established Difference in Pathogenicity between strains

- Isolates of *T. b. rhodesiense* from South-Eastern Uganda
- Collected 1989-1992
- Patients reporting showed two broad clinical histories
- Virulence replicated in mice

Course of infection	Acute Mean 4 wks	Chronic Mean 20 wks
Stage of disease at first presentation	Early	Late
Chancre	92% of cases	None recalled
Bloodstream Parasitaemia	+++	-
Trypanosomes in CSF	-	+
WBC in CSF	< 5 cells / mm ³	> 5 cells / mm ³
Zymodeme	<i>Busoga</i> (predominantly <i>B17</i>)	<i>Zambesi</i> (predominantly <i>Z310</i>)

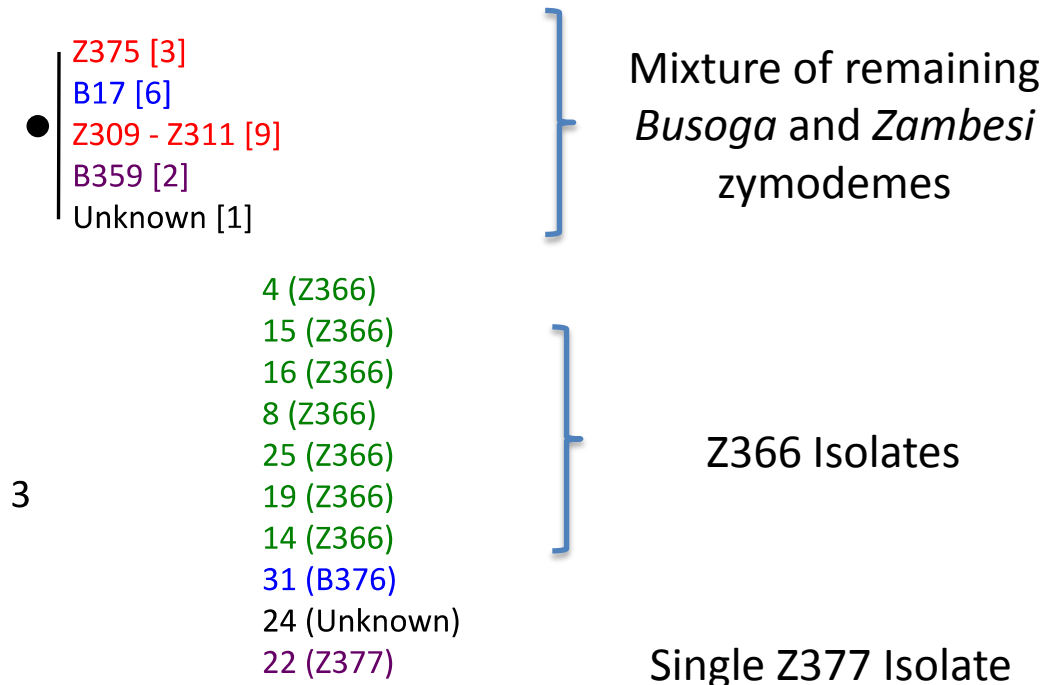
Established Difference in Pathogenicity between strains



- Survival in very susceptible A/J mice corresponds to virulence in humans

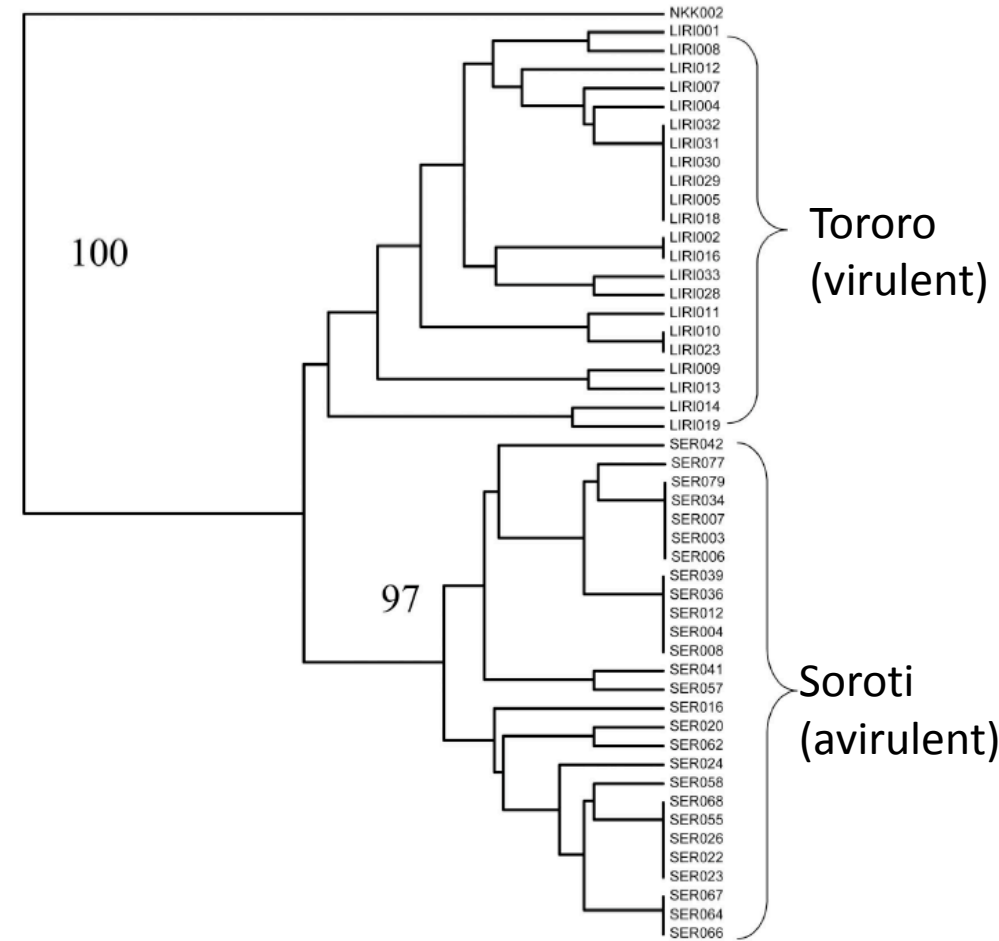
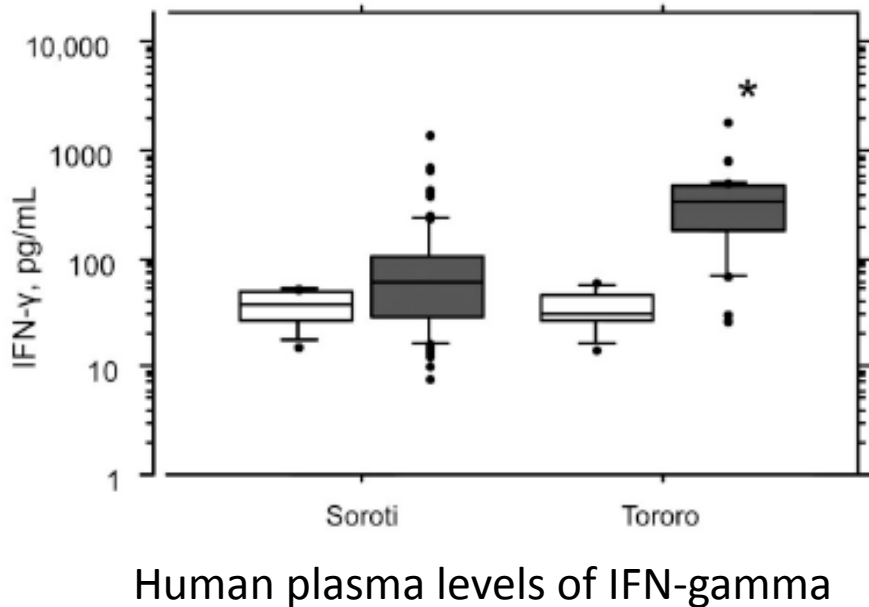
Multilocus genotyping fails to distinguish Z310 and B17 zymodeme isolates

- 31 isolates genotyped at 11 informative microsatellite loci
- Difficult to distinguish Zambesi 310 and Busoga 17 populations
- Strain with intermediate phenotype – Z366 - is distinguishable using the same markers
 - All Z366 isolates are from a specific 4-year outbreak in Bugiri region in 1993



Bootstrapped Dendrogram
(UPGMA; Jaccard's similarity index)

Spatially and Genetically Distinct African Trypanosome Virulence Variants Defined by Host Interferon-g Response (2002-2004 outbreak)



Dendrogram showing similarities between trypanosome multilocus microsatellite genotypes.

Hypothesis

- Established clear differences in phenotype in both man and in experimental infections in mice
- Microsatellite genotyping shows that Z310 and B17 are indistinguishable using the eleven markers tested
- Can “next-generation” DNA sequencing provide some clues as to what differences there are between them?

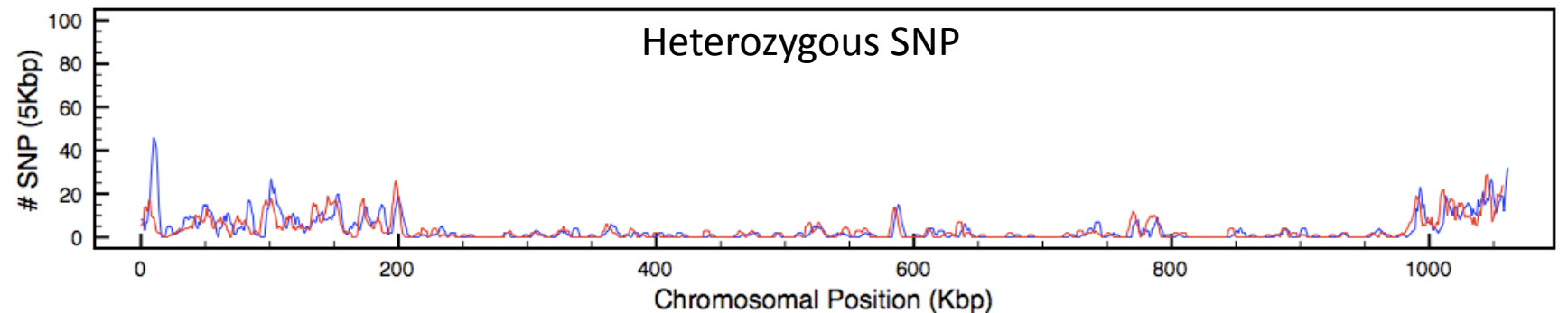
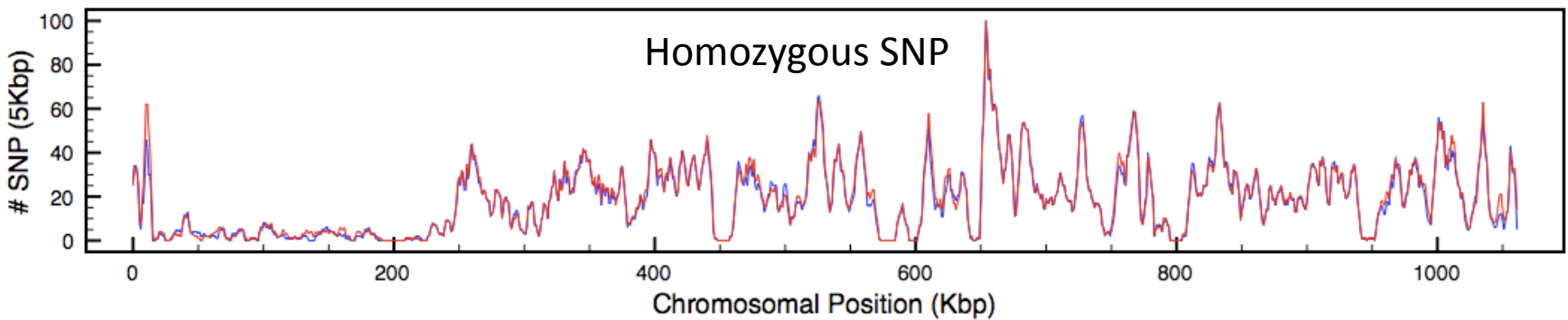
SOLiD Sequencing: Single Nucleotide Polymorphism (SNP) discovery

- Sequenced one isolate each of *Z310* and *B17*
 - Life Technologies SOLiD (v3) platform
 - Single run generated >60X of both genomes!
- Reads aligned against *T. b. brucei* reference genome
- Genomes >97% similar
 - Z310 (less virulent)*
 - 13,716 polymorphisms
 - B17 (more virulent)*
 - 18,420 polymorphisms



SOLiD sequencing reveals distinct patterns of SNP distribution

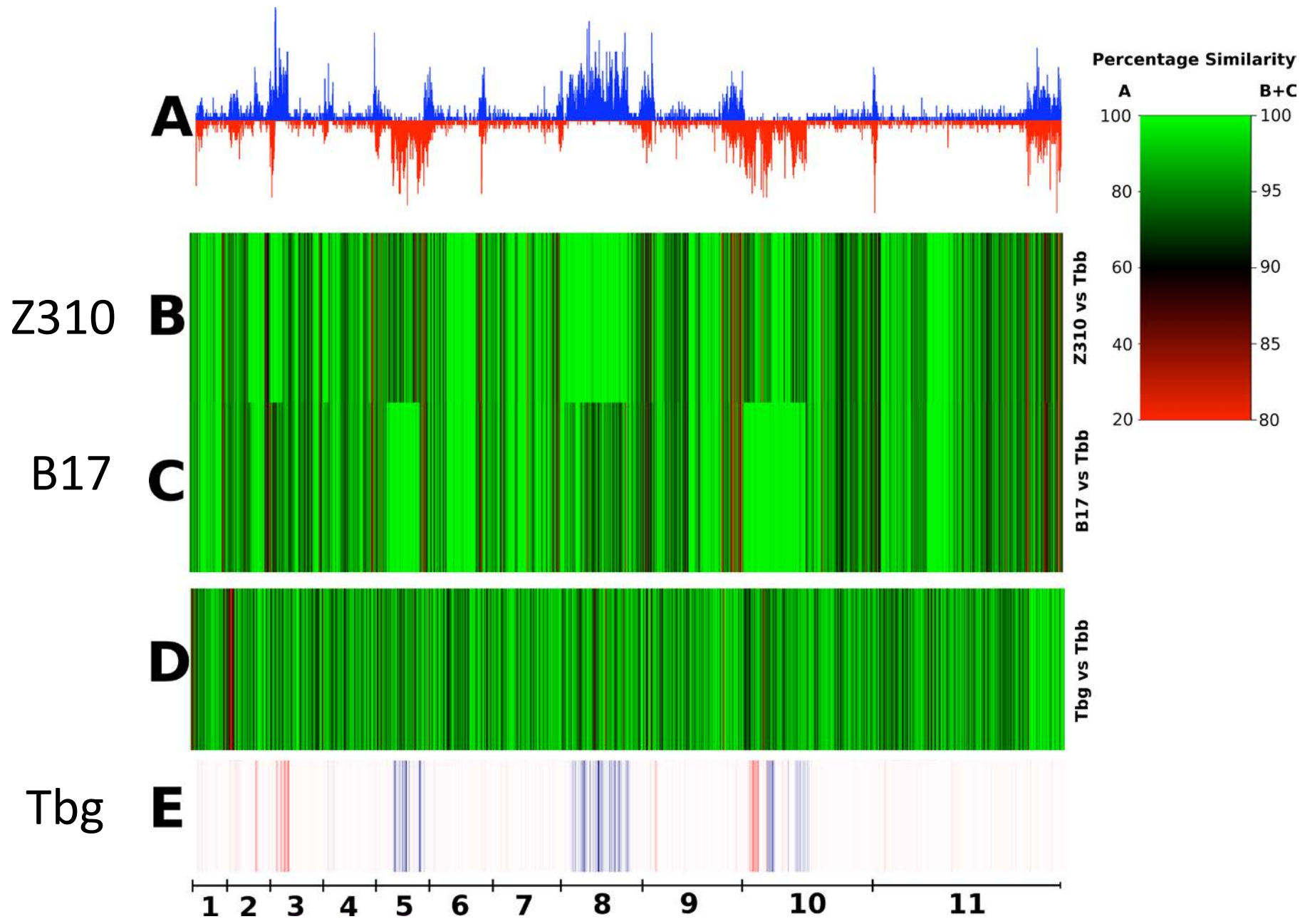
- Plot of 5Kbp 'moving average' of raw SNP (vs. *T. b. brucei* sequence) across each chromosome revealed 'regions' of loss of heterozygosity



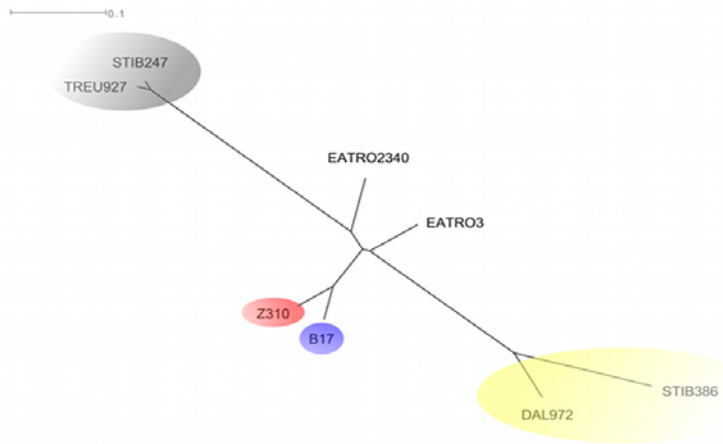
Heterozygous region
(both zymodemes)

Homozygous region

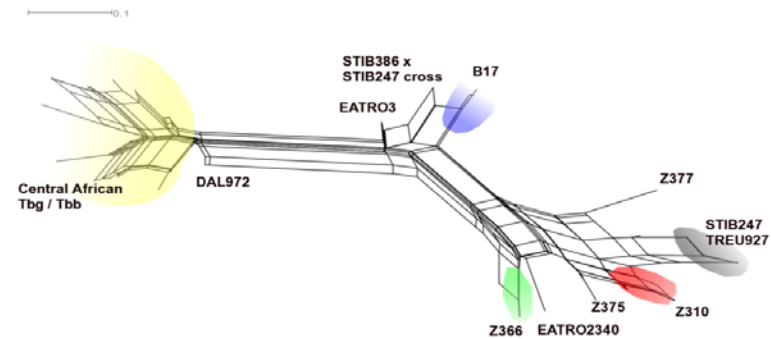
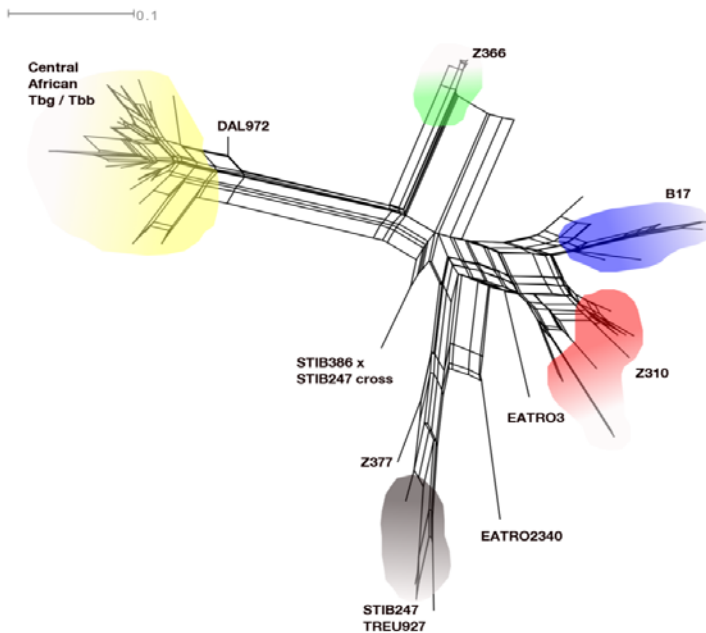
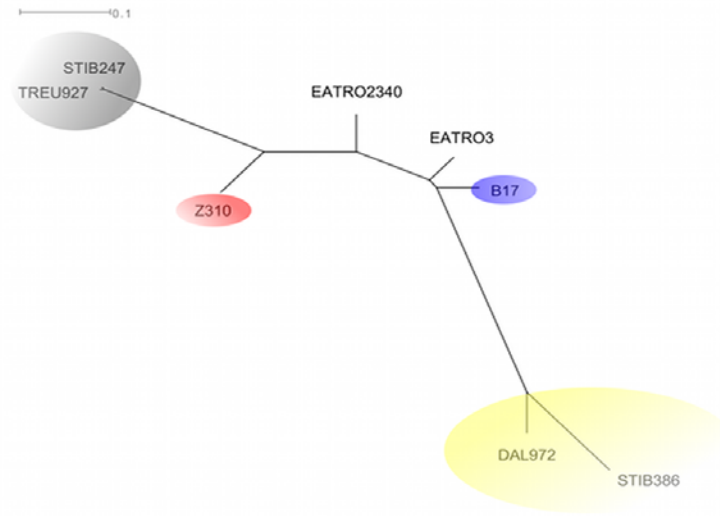
— Z310 (less virulent)
— B17 (more virulent)



Whole Genome



Chromosome 8



Virulent strain (B17)

Less-virulent strain (Z310)

Trypanosomiasis: “Sleeping Sickness” – Human Disease

Affects 500,000 annually across 36 countries

Caused by two species of *T. brucei* parasite:

T. brucei gambiense

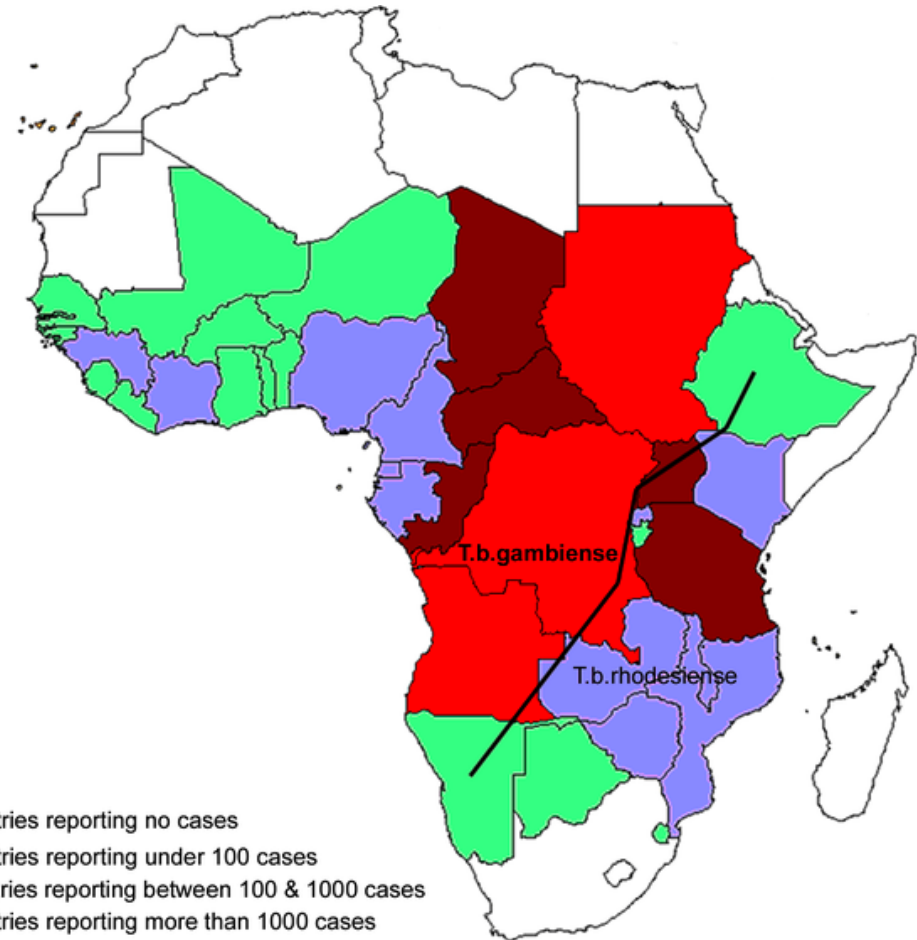
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T. brucei brucei

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Identification of possible marker for differences in virulence

- Isocitrate dehydrogenase is the only MLEE enzyme that distinguishes *Zambesi* from *Busoga* isolates
- Three heterozygous non-synonymous SNP predicted within Chromosome 8 copy of ICD gene in SOLiD sequenced *B17* isolate
 - Confirmed by Sanger sequencing on PCR on bloodstream stabilates
 - Found to correlate with zymodeme as per MLEE pattern
- Actually two copies of ICD, None found in chromosome 11 copy of ICD

B17
Z310



Alignment of amino acid sequence of the beginning (left) and end (right) of the chromosome 8 copy of Isocitrate dehydrogenase gene

Conclusions

- Strains of *T. b. rhodesiense* have a reproducible difference in virulence phenotype
- Microsatellite data suggests either isolates are very closely related, and/or not sensitive enough to detect variation
- Next generation sequence data reveal high level of genomic similarity between strains (>97%)
- Identified discrete regions of heterozygosity which are different between the two strains that isn't detected by other methods
- Polymorphism(s) on ~80% of chromosome 8 differ between strains may help regulate differences in virulence between *Busoga* and *Zambesi* strains

Trypanosomiasis:

“Nagana” – Animal Disease

- Caused largely by *T. congolense* and *T. vivax*
- Symptoms in domestic animals include fever, muscle wastage and anaemia.
- Affects over ten million Km²
- 30% African cattle are at risk of infection
- Losses in livestock and crop production amount to more than ~\$1 billion per annum



Host Genetics: Trypanotolerance

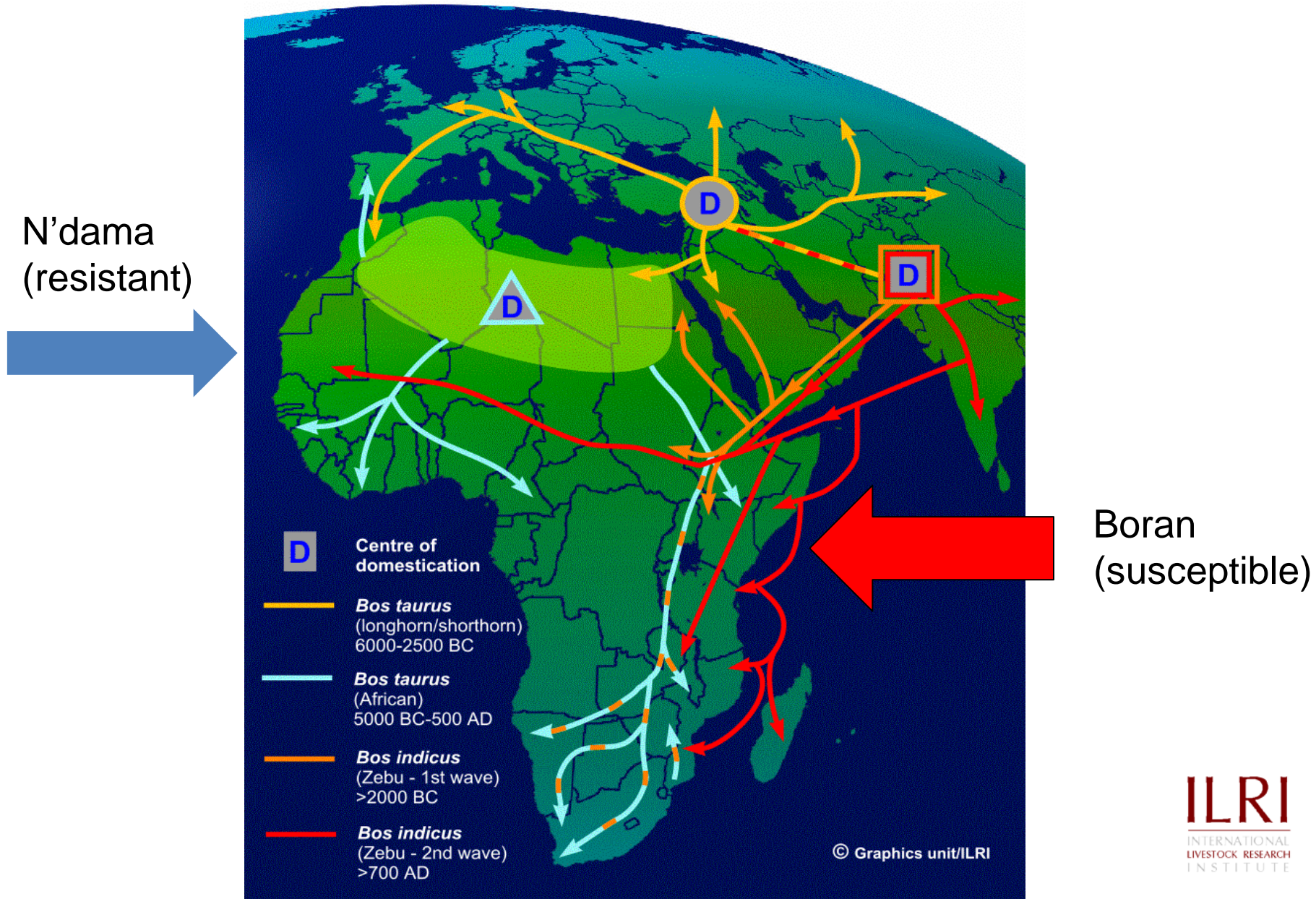


Resistant: N'dama
(*Bos taurus*)

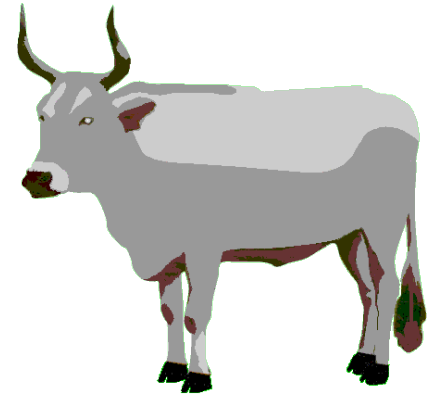


Susceptible: Boran
(*Bos indicus*)

Origin and migration routes of domestic cattle in Africa

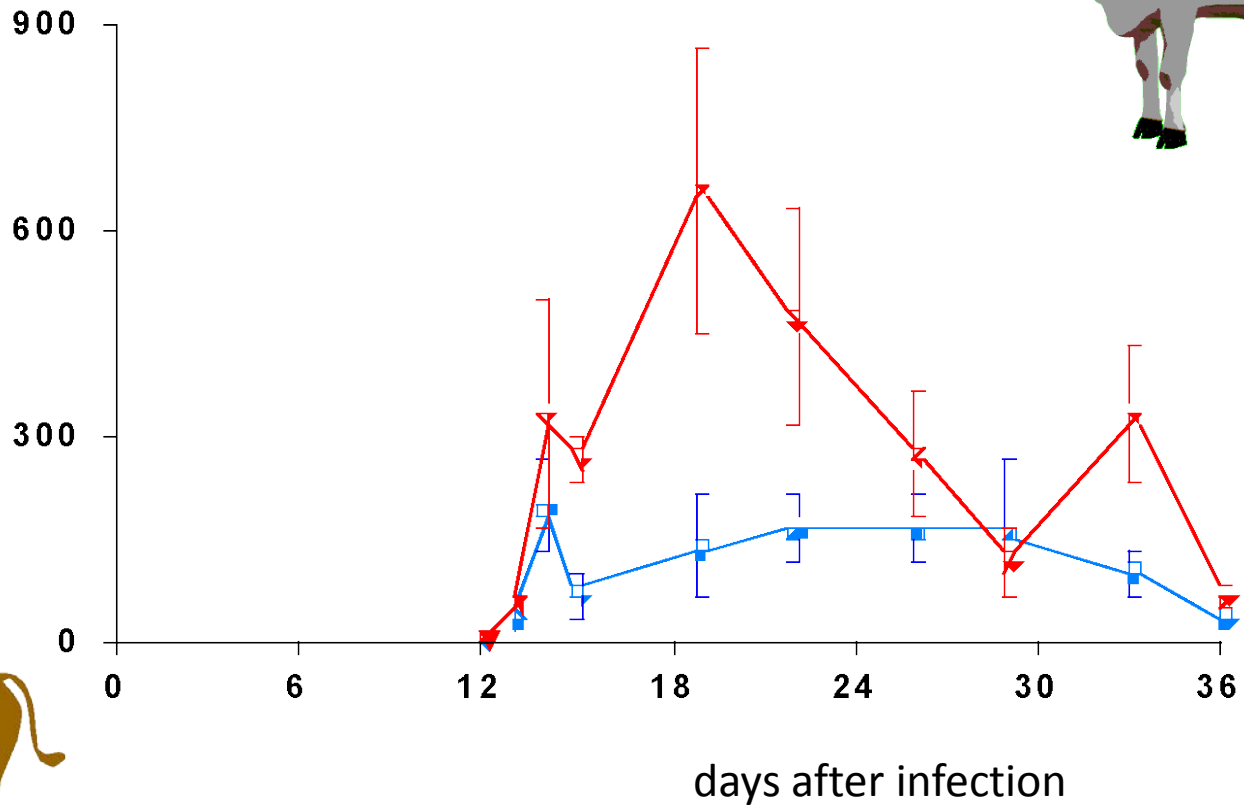


Parasitaemia

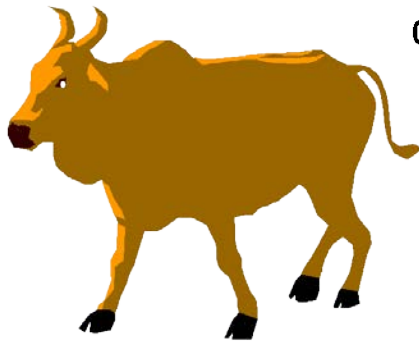


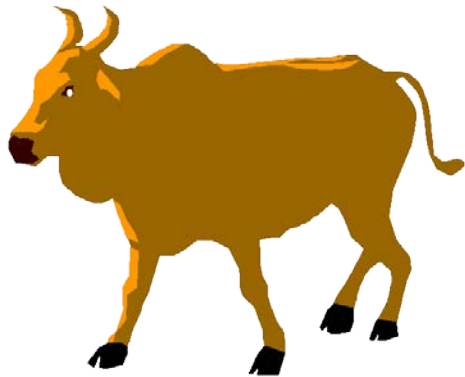
N'Dama

1st wave
parasitemia



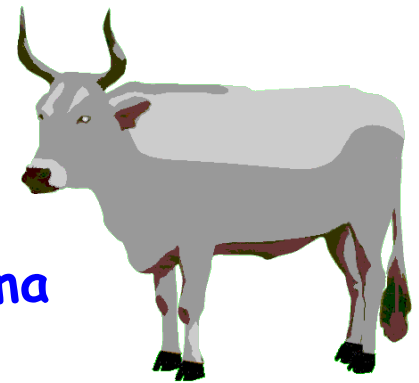
Boran





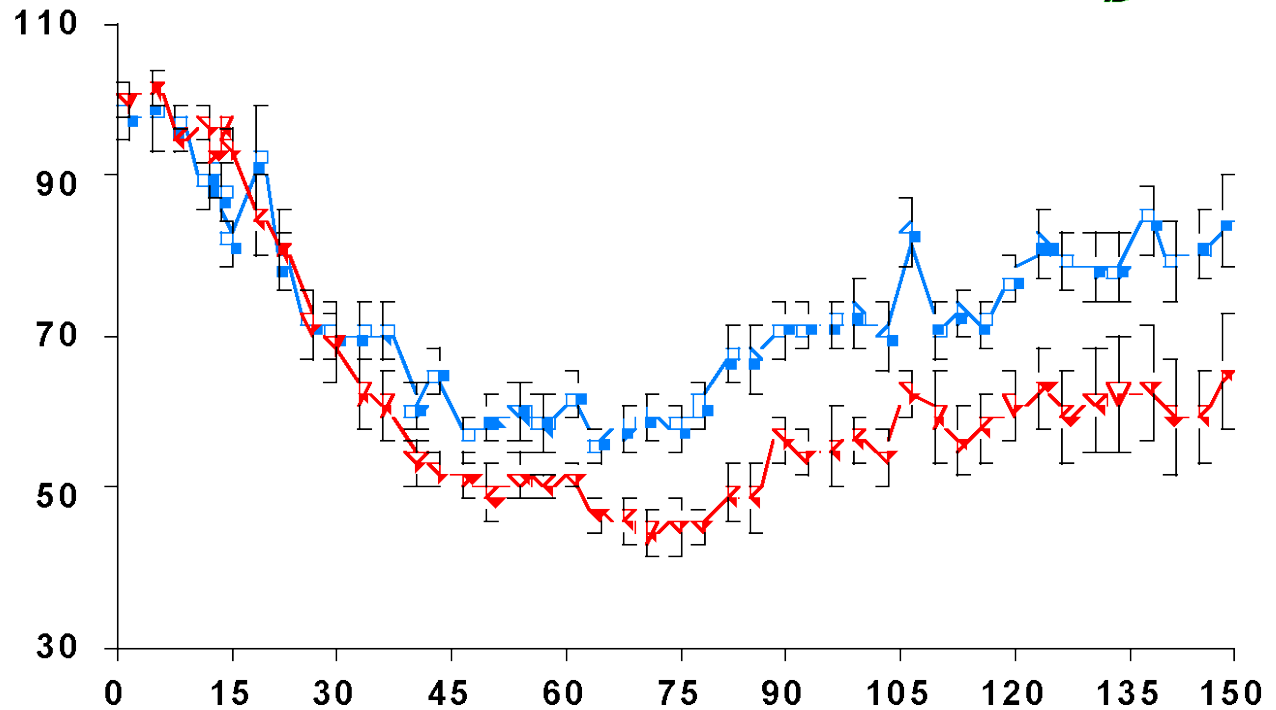
Boran

Anaemia (PCV)



N'Dama

% change in PCV



days after infection

Host Genetics: The mouse model



N'dama
(*Bos taurus*)

Resistant:



C57BL/6 (Black Six) mice



Boran
(*Bos indicus*)

Susceptible:



A/J and BALB/c mice

Tolerance to *T. congolense* infection: Mapping in Inbred Mice

- Tolerance in the mouse model has a major genetic component
- Mapping in genetic crosses has revealed three regions that control resistance / susceptibility

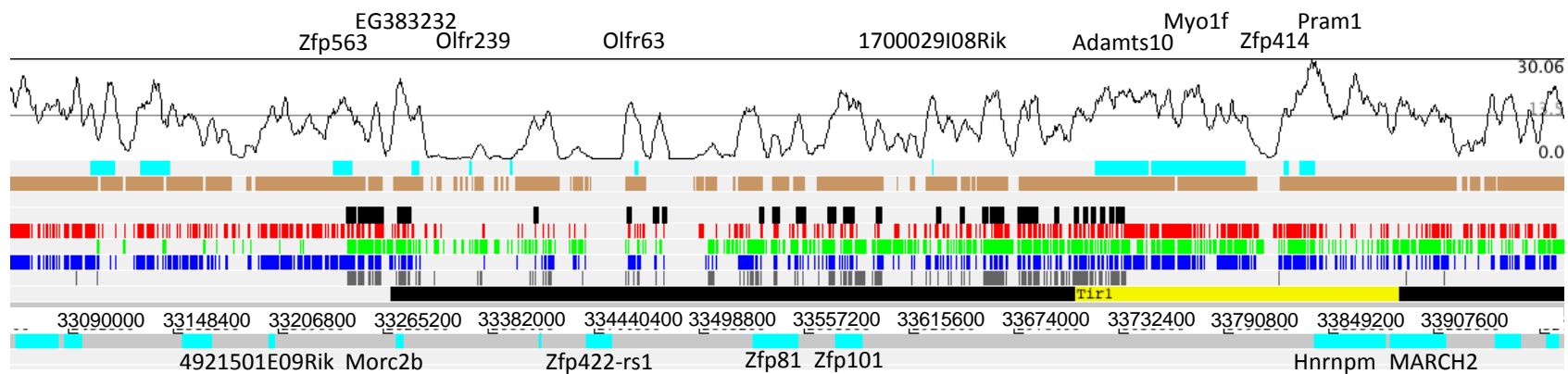
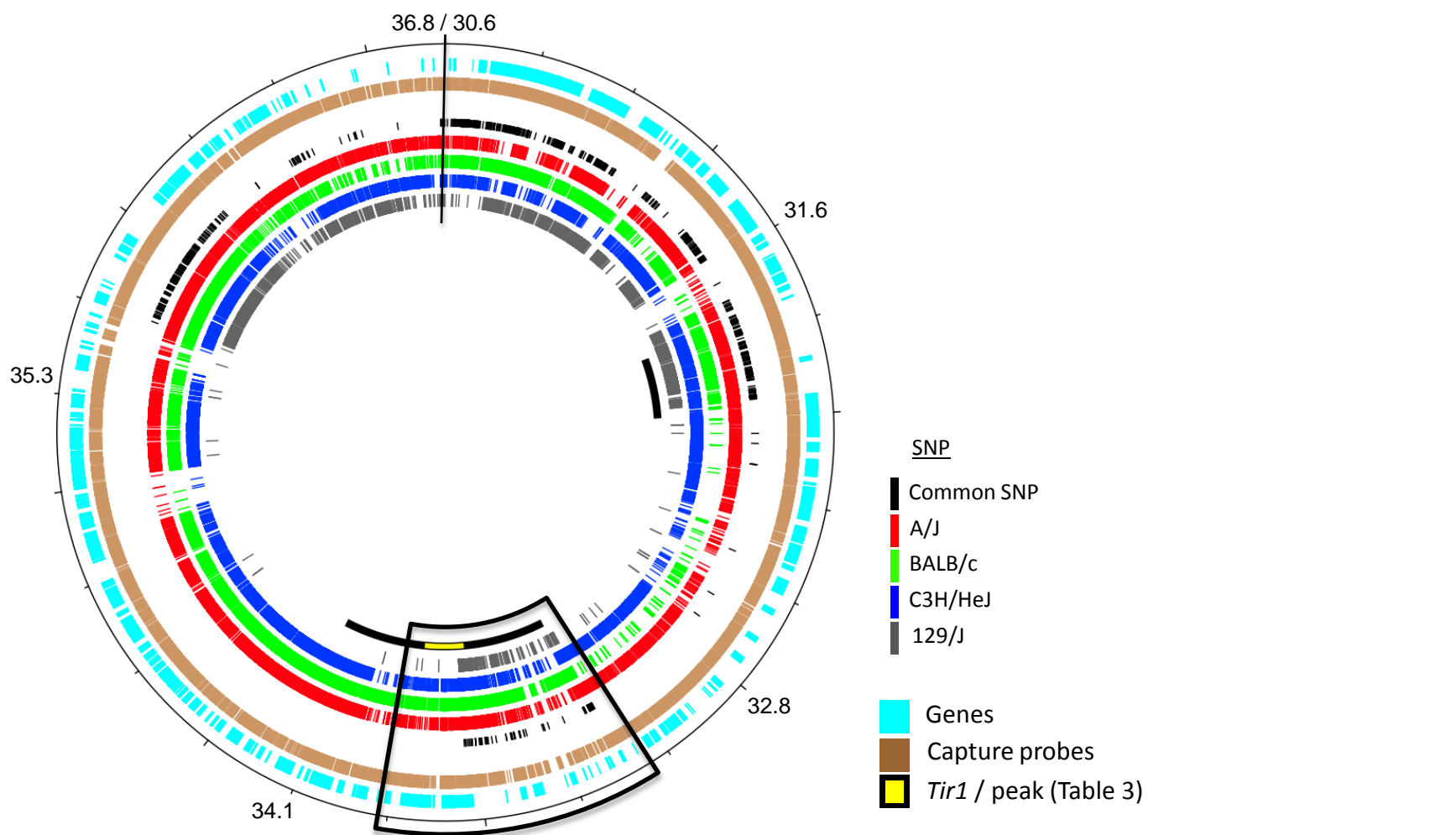
Quantitative trait loci (QTL)

- Chromosome 17 – *Tir1*
- Chromosome 5 – *Tir2*
- Chromosome 1 – *Tir3a-c*
- *Problem – these regions contain hundreds of genes that may be controlling the response to infection*

Sequence Capture of Inbred Mice

- “Capture” region of interest (i.e. *Tir1*)
- Resequence 4 susceptible breeds of mice
- Identify “shared” polymorphisms

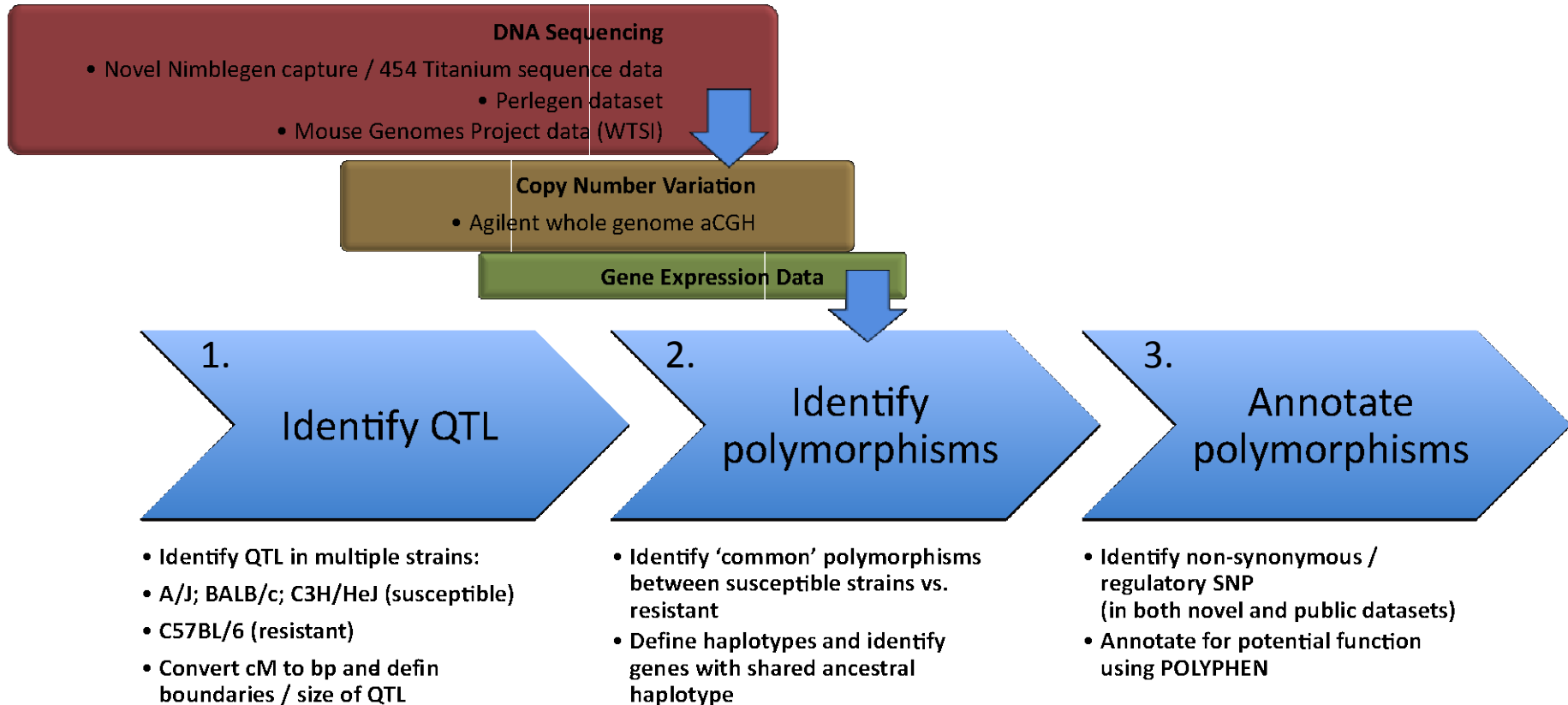




Targeted Resequencing of *Tir1*:

- Identified one candidate gene with two functional SNP
- One SNP is predicted to affect the function of the protein
 - *Pram1* - PML-retinoic acid receptor alpha regulated adaptor molecule 1
 - Predicted to have an effect on the early- pro-inflammatory response after infection

Utilising Genomic Technologies to reduce the number of candidate genes affecting Trypanotolerance in mice



Trypanotolerance:

Number of “candidate genes” under QTL

QTL	Before	After
<i>Tir1</i>	43	27
<i>Tir2</i>	210	14 – 74
<i>Tir3a-c</i>	20 - 650	10 – 355*

* Candidate region revealed later in pipeline

Trypanotolerance Summary:

- Attempted to characterise every Single Nucleotide Polymorphism in a 4.2Mbp region of 4 breeds of inbred mouse.
- Systematically reduced number of candidate genes at *Tir1* from 73 to just 7.
- Functional testing of identified pathways in cattle are now feasible (and underway)

Summary (2)

- Early sequencing work established ‘reference’ genomes on which modern NGS techniques are now based.
- Cheaper, quicker and easier* to produce large numbers of whole genome sequences rapidly to compare
 - Pathogen
 - Host

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